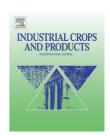


available at www.sciencedirect.com







Short communication

Ultrahigh CO₂ levels enhances cuphea growth and morphogenesis[☆]

Brent Tisserat*, Steven F. Vaughn, Mark A. Berhow

New Crops and Processing Technology Research, National Center for Agricultural Utilization Research, Agricultural Research Service-USDA, 1815 N. University St., Peoria, IL 61604, USA

ARTICLE INFO

Article history:
Received 25 April 2007
Received in revised form
25 June 2007
Accepted 26 June 2007

Keywords: Cuphea CO₂ enrichment Rooting Seedling dormancy Seedling vigor

ABSTRACT

Applications of ultrahigh CO_2 treatments accelerated cuphea (*Cuphea viscosissima* × C. lance-olata 'PSR23') growth and development and aided in seedling establishment. The growth (fresh weight) and morphogenesis (number of leaves and roots and seedling length) were determined in cuphea seedlings exposed to 350, 1500, 3000, 10,000, or 30,000 μ mol mol $^{-1}$ CO_2 for 30 days under greenhouse conditions. Greater CO_2 levels, especially the ultrahigh levels (i.e. \geq 3000 μ mol mol $^{-1}$ CO_2) resulted in significantly higher ($P \leq 0.05$) fresh weights, leaf numbers, root numbers, and seedling lengths compared to seedlings grown under ambient air (350 μ mol mol $^{-1}$ CO_2). For example, cuphea 'PSR23' Morris heavy seedlings showed the greatest seedling fresh weight, leaf number, root number, and seedling length when supplemented with 10,000 μ mol mol $^{-1}$ CO_2 increasing 607%, 184%, 784%, and 175%, respectively, when compared to seedlings grown without CO_2 enrichment.

Published by Elsevier B.V.

1. Introduction

Cuphea (Lythraceae) is an annual native species which produces an oil seed that is high in saturated, medium-chain triacylglycerols and can be used for making detergents, surfactants, lubricants, and other products (Cermak et al., 2005; Phippen et al., 2006). The unique seed oil composition coupled to high commercial demand suggests the possibility for future cuphea cultivation as a new crop. However, a number of negative traits retard optimum cuphea cultivation such as indeterminate flowering, seed shattering, seed dormancy, lack of self-pollination, and occurrence of sticky glandular hairs on foliage (Knapp, 1993; Isbell,

2002). Cuphea 'PSR23' seedlings unique among cuphea lines expresses temporary arrested growth following germination which may last from 2 to 5 weeks and hampers rapid breeding efforts. Breeding work would benefit from accelerated seedling growth, therefore, we sought to test the elevated levels of CO₂ on cuphea. Further, methods to accelerate cuphea seedling growth would also benefit tissue culture plantlet establishment in the soil and their subsequent growth. Prior studies with elevated CO₂ environments have demonstrated its ability to enhance growth with mint (Tisserat, 2002), pine (Groninger et al., 1996; Tisserat and Vaughn, 2003), potato (Schapendonk et al., 2000), soybean (Rodgers et al., 2004), and sweetgum (Groninger et al., 1996). The influence of ele-

^{*} Names are necessary to report factually on available data; however, the USDA neither guarantees nor warrants the standard of the product, and the use of the name by USDA implies no approval of the product to the exclusion of others that may also be suitable.

^{*} Corresponding author. Tel.: +1 309 6816289; fax: +1 309 6816524.

vated CO_2 treatments on cuphea has not been determined to date

Usually, CO_2 enrichment (700–1000 μ mol mol $^{-1}$ CO_2) promotes enhanced growth of seedlings in several different species (Groninger et al., 1996; Schapendonk et al., 2000; Rodgers et al., 2004). However, it has been recently demonstrated that ultrahigh CO_2 levels (\geq 3000 μ mol mol $^{-1}$ CO_2) enhances growth in several C-3 photosynthesis species more significantly than employing lower CO_2 levels (Tisserat, 2002; Tisserat and Vaughn, 2003). The present study was conducted to determine the growth and morphogenesis response of immature cuphea seedlings to a broad range of CO_2 levels and to determine if short CO_2 treatments may reduce seedling nursery growing time.

2. Materials and methods

2.1. CO₂ flow systems

CO2 flow through testing chambers were made out of a transparent polycarbonate box and lid (Consolidated Plastics, Twinsburg, OH) (45.7 cm $W \times 66$ cm $L \times 76.2$ cm D; 162.6 L capacity) with a silicone tape gasket (Furon, New Haven, CT) attached. Three polypropylene spigots (Ark-Plas Products, Flippin, AR) attached to 0.45 µm air vents (Gelman Science, Ann Arbor, MI) were mounted to the box. The box and lid were clamped with 12 equally spaced stationary binding clips (50 mm L) to create an air tight seal. CO2 was provided by a gas cylinder (BOC Gases, Edison, NJ) rated 99.8% pure and was mixed with an ambient air flow produced by an aquarium air pump (Whisper 2000, Carolina Biological Supply Company, Burlington, NC) via a flow meter (Cole Parmer Instrument Co., Niles, IL) to provide 350 (control, without CO₂ enrichment), 1500, 3000, 10,000, or $30,000 \,\mu \text{mol mol}^{-1}$ CO₂. CO₂ ranges \geq 10,000 μ mol mol⁻¹ CO₂ were adjusted using a LIRA infrared gas analyzer (model #3000, Mine Safety Appliances Company, Pittsburgh, PA) and CO_2 ranges $\leq 3000 \,\mu\text{mol}\,\text{mol}^{-1}$ CO_2 were adjusted with a Li-Cor CO2/H2O infrared gas analyzer (model LI-6262, Li-Cor, Inc., Lincoln, NE). The CO2 and air streams were added at 2000 mL min⁻¹ during the photoperiod (14 h/day). Control seedlings were given a stream of ambient air generated by the aquarium pump only. No CO2 or air control was applied during the dark (10 h/day).

2.2. Experiments

2.2.1. Cuphea viscosissima × C. lanceolata L.

'PSR23' types 'McCoy GT #1', 'Morton GT #1' and 'Morris heavy' seedlings were grown in cone-tainers (Hummert Internat., Earth City, MO; 25 mm in diameter \times 160 mm in length) containing 10 g of a soilless medium formulated with peat moss:vermiculite mixed at 1:1 volume ratio and amended with 10.9 g kg $^{-1}$ Micromax (Scotts Co., Marysville, OH) and 62.3 g kg $^{-1}$ Osmocote 14-14-14 (Scotts Co.). To determine the optimum CO $_2$ levels for growth and morphogenesis, forty 2-week-old uniformed sized cuphea seedlings were grown under 350, 1500, 3000, 10,000, or 30,000 μ mol mol $^{-1}$ CO $_2$ within 162.2 L transparent containers. Seedlings were watered two times per week and not additionally fertilized during the

experimental CO_2 incubation periods. Seedlings were grown within CO_2 test chambers were maintained in a temperature-controlled greenhouse. All treatments were run concurrently and exposed to the same temperature and lighting regimes during the experiment. Experiments were repeated twice during March through June 2006. Average daily temperature was 25.2 °C and varied from 20.8 to 29.2 °C. Illumination during experiments was provided by natural sunlight with an average daily photosynthetic photon flux of 550 μ mol m⁻² s⁻¹. After 30 days of incubation, data on whole seedling fresh weight, leaf number per seedling, root number per seedling, and length per seedling were recorded from five randomly selected seedlings for each CO_2 treatment. Best fit regression equations were calculated (Table Curve.2D ver. 5.0, 2000 AISN Software, Inc.) for each response variable as a function of CO_2 treatment. Regres-

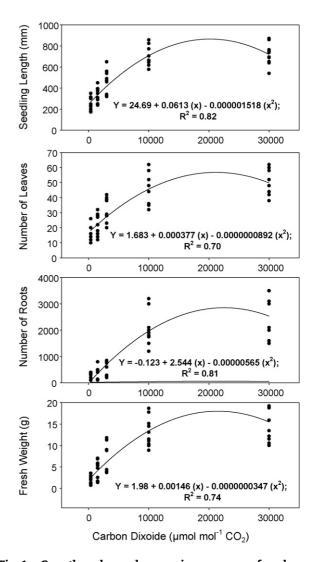


Fig. 1 – Growth and morphogenesis responses of cuphea 'PSR23' Morris heavy seedlings exposed to various concentrations of CO_2 for 30 days. Observations from two replications are presented. Regression coefficients of determination (R^2) and regression equations between CO_2 and length/seedling, leaves/seedling, roots/seedling, and fresh weight/seedling are given. All repressions were significant at $P \leq 0.05$.

Table 1 - Pearson correlation coefficients values for				
growth, morphogenesis and CO ₂ ^a				

	CO ₂	Leaves	Roots	Fresh weight (g)
CO ₂	_	0.743	0.816	0.755
Leaves	0.743	-	0.825	0.927
Roots	0.816	0.825	_	0.878
Fresh weight (g)	0.755	0.927	0.878	-
Seedling length (cm)	0.743	0.854	0.818	0.864

^a All values were significant at $P \le 0.05$. Observations: leaves, 10; roots, 10; fresh weight, 10; seedling length, 10.

sion model analyses and 95% confidence limits on the mean predicted values of the response variables from the individual equations were obtained from the REG procedure of SAS (SAS Institute, ver. 8.2, Cary, NC).

3. Results and discussion

Short-term exposure (30 days) to various elevated CO2 levels significantly affected growth and morphogenesis of seedlings for all cuphea ecotypes tested (Fig. 1). Increasing CO2 concentrations up to 10,000 µmol mol⁻¹ CO₂ proportionally increased whole seedling fresh weight, root number, seedling length and leaf number compared to seedlings grown under ambient CO2 levels. Smaller increases in growth and morphogenesis responses occurred in cuphea seedlings when grown under 30,000 µmol mol⁻¹ CO₂ compared to that obtained employing $10,000 \,\mu\text{mol}\,\text{mol}^{-1}\,\text{CO}_2$. Fresh weight of seedlings, leaves per seedling, roots per seedling, and seedling length in cuphea Morris heavy seedlings increased 607%, 184%, 784%, and 175%, respectively, after 30-day exposure to 10,000 μmol mol⁻¹ CO₂ over those obtained from seedlings grown on ambient CO2 levels (Fig. 1). All cuphea seed types showed similarly response trends to increasing CO2 concentrations (data not shown). Other investigators have found that elevated CO2 environments (e.g., $600-800 \,\mu\text{mol mol}^{-1} \,\text{CO}_2$) enhanced other species fresh weights (Groninger et al., 1996; Schapendonk et al., 2000). For example, Groninger et al. (1996) found a 37% biomass increase in loblolly pine seedling when cultured under $806 \,\mu mol \, mol^{-1} \, CO_2$ after several months of exposure. In these 30-day experiments, we obtained a significant increase in biomass employing ultrahigh CO_2 ($\geq 3000 \,\mu\text{mol mol}^{-1} \,CO_2$) levels over ambient air. There are close associations between CO2 levels, growth and morphogenesis responses (Table 1). As CO₂ levels increase, growth and morphogenesis responses correspondingly increase. CO2 levels had strong significant positive correlations with fresh weights, seedling length, number of leaves and number of roots. Similarly, all growth, morphogenesis responses and CO_2 levels showed strong significant positive correlations among themselves.

Our studies were conducted for relatively short periods, i.e. 30 days, because we sought to develop a suitable short-term nursery CO_2 application treatment which would be financially and spatially practical for growing cuphea seedlings in ultrahigh CO_2 conditions within a greenhouse environment. We noted that significant differences in seedling growth were visible after 7–10 days. Ten-day-old seedlings grown in 10,000 μ mol mol $^{-1}$ CO_2 were considerably larger than seedlings grown under ambient air. Plants removed from CO_2 chambers continued to grow normally thereafter and did not exhibit any adverse effects from their prior CO_2 treatments. Our data suggests that ultrahigh CO_2 treatments may be effective for enhancing cuphea growth and benefit breeding treatments.

REFERENCES

Cermak, S.C., Isbell, T.A., Isbell, J.E., Akerman, G.G., Lowery, B.A., Deppe, A.B., 2005. Batch drying of cuphea seeds. Ind. Crops Prod. 21, 354–459.

Groninger, J.W., Seiler, J.R., Zedaker, S.M., Berrang, P.C., 1996. Photosynthetic response of loblolly pine and sweetgum seedling stands to elevated carbon dioxide, water stress, and nitrogen level. Can. J. For. Res. 26, 95–102.

Isbell, T.A., 2002. Current progress in the development of *Cuphea*. Lipid Technol. 14, 77–80.

Knapp, S.J., 1993. Breakthroughs towards the domestication of Cuphea. In: Janick, J., Simon, J.E. (Eds.), New Crops. Wiley, New York, pp. 372–379.

Phippen, W.B., Isbell, T.A., Phippen, M.E., 2006. Total seed oil and fatty acid methyl ester contents of Cuphea accessions. Ind. Crops Prod. 24, 52–59.

Rodgers, A., Allen, D.J., Davey, P.A., Morgan, P.B., Ainswoth, E.A., Bernacchi, C.J., Cornic, G., Dermody, O., Dohleman, F.G., Heaton, E.A., Mahoney, J., Zhu, E.H., Delucia, X.-G., Ort, D.R., Long, P.S., 2004. Leaf photosynthesis and carbohydrate dynamics of soybeans grown throughout their life-cycle under free-air carbon dioxide enrichment. Plant Cell Environ. 27, 449–458.

Schapendonk, A.H.C.M., van Oijen, M., Dijkstra, P., Pot, C.S., Jordi, W.J.R.M., Stoopen, G.M., 2000. Effects of elevated CO₂ concentration on photosynthetic acclimation and productivity of two potato cultivars grown in open-top chambers. Aust. J. Plant Physiol. 27, 1119–1130.

Tisserat, B., 2002. Influence of ultrahigh carbon dioxide levels on growth and morphogenesis of Lamiaceae species in soil. J. Herbs Spices Med. Plants 9, 81–89.

Tisserat, B., Vaughn, S.F., 2003. Ultrahigh CO_2 levels enhance loblolly pine seedling growth, morphogenesis and secondary metabolism. HortScience 38, 1083–1085.